Stereochemistry of the Reduction of Nicotinic Acid when It Serves as a Precursor of Anatabine

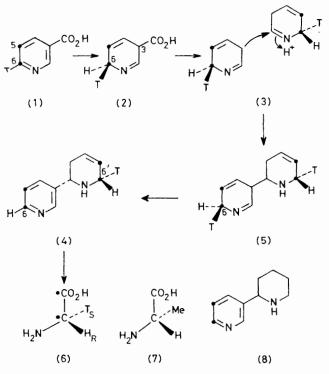
By Edward Leete

(Natural Products Laboratory, School of Chemistry, University of Minnesota, Minneapolis, Minnesota 55455)

Summary Anatabine derived from $[5,6^{-14}C,1^{3}C,6^{-3}H]$ nicotinic acid (which is a precursor of both rings of the alkaloid) was almost devoid of tritium in its pyridine nucleus, but retained 100% at C-6'; this tritium was in a *pro-S* position, indicating that nicotinic acid was reduced at C-6 by the introduction of a hydrogen in the *pro-R* position.

We have previously established that both rings of (-)anatabine (4) are derived from nicotinic acid,^{1,2} and we have suggested that its biosynthesis, illustrated in the Scheme, involves the dimerization of 2,5-dihydropyridine (3), which is formed by the decarboxylation of the hypothetical intermediate, 3,6-dihydronicotinic acid (2). We considered that the stereochemistry of the reduction which occurs at C-6 of nicotinic acid could be probed by determination of the chirality at C-6' of the anatabine which would be formed from $[6^{-3}H]$ nicotinic acid.

Accordingly, a mixture of $[5,6^{-14}C,1^{3}C_{2}]$ nicotinic acid² and $[6^{-3}H]$ nicotinic acid³ (³H/¹⁴C, 7·0) was fed, by the wick method, to *Nicotiana glauca* plants for seven days. The resultant anatabine was labelled equally in both rings with ¹³C (determined by ¹³C n.m.r. spectroscopy²) and had ¹⁴C activity: $1\cdot91 \times 10^{7}$ d.p.m. mmol⁻¹ ($3\cdot8\%$ specific incorporation), ³H/¹⁴C, $3\cdot7$ (53% retention of tritium). It was converted into *N*-benzoylanatabine which was oxidized with acidic potassium permanganate as previously described¹ to yield hippuric acid ($^{14}C: 9\cdot6 \times 10^{7}$ d.p.m. mmol⁻¹, ³H/¹⁴C, $6\cdot95, 100\%$ retention of tritium) and nicotinic acid



SCHEME. (Heavy dots indicate position of ¹⁴C and ¹³C).

(14C: 9.7×10^7 d.p.m. mmol⁻¹, ${}^{3}H/{}^{14}C$, 0.52, 93% loss of tritium). The loss of tritium from C-6 of the pyridine ring of anatabine is in accord with previous work on the incorporation of [6-3H]nicotinic acid into nicotine.^{3,4} We have rationalized this loss by suggesting² that the final step in the biosynthesis of nicotine involves the stereospecific loss of the hydrogen which was originally present at C-6 of nicotinic acid. We presume that the final step in the biosynthesis of anatabine, namely the dehydrogenation of (5), is also stereospecific involving the loss of tritium from C-6. Anabasine (8), the major alkaloid of N. glauca was also isolated (14C: 7.33×10^6 d.p.m. mmol⁻¹, 1.98% specific incorporation), and had ³H/¹⁴C, 0·37, representing a 95% loss of tritium from C-6.

The hippuric acid obtained from anatabine was hydrolysed with hydrochloric acid to afford glycine $({}^{3}H/{}^{14}C, 7 \cdot 0)$. Control experiments with [2-14C,3H]hippuric acid indicated that its ³H/¹⁴C ratio remained unchanged in the course of the permanganate oxidation and subsequent hydrolysis to glycine. The chirality of the glycine was determined enzymatically.⁵ When L-glutamic-pyruvic transaminase

(E.C. No 2.6.1.2) which normally catalyses the reversible transfer of an amino group from L-alanine to 2-oxoglutarate, acts upon glycine in the absence of an amino group acceptor it is the pro-R hydrogen at C-2 of glycine, corresponding to the α -hydrogen of L-alanine (7) which is labilized. When the glycine derived from anatabine was incubated with this enzyme there was no loss of tritium (${}^{3}H/{}^{14}C$, $6 \cdot 9 \pm 0 \cdot 1$). Non-chiral [2-14C,3H]glycine incubated with this enzyme under identical conditions lost $50 \pm 2\%$ of its tritium relative to the ¹⁴C. This enzyme assay was carried out in triplicate. This result indicates that the glycine derived from anatabine was labelled with tritium in the pro-S position, as illustrated in structure (6). One can thus deduce that the initial reduction of nicotinic acid occurs by introduction of a hydrogen at C-6 in the pro-R position as indicated in structure (2).

This investigation was supported by a research grant from the National Institutes of Health, U.S. Public Health Service.

(Received, 17th April 1978; Com. 396.)

- ¹ E. Leete and S. A. Slattery, J. Amer. Chem. Soc., 1976, 98, 6326. ² E. Leete, Bioorganic Chem., 1977, 6, 273.
- ³ R. F. Dawson, D. R. Christman, A. D'Adamo, M. L. Solt, and A. P. Wolf, J. Amer. Chem. Soc., 1960, 82, 2628. ⁴ E. Leete and Y-Y. Liu, Phytochemistry, 1973, 12, 593.
- ⁵ P. Besmer and D. Arigoni, Chimia (Switz), 1968, 22, 494.